Somatic Embryogenesis in *Abies fraseri* (Fraser fir):

Optimizing Levels of Abscisic Acid, Polyethylene Glycol & Maltose for Maturation

Robert Thomas, Lilian Matallana and John Frampton Department of Forestry and Environmental Resources North Carolina State University, Raleigh NC

What is Embryogenesis?

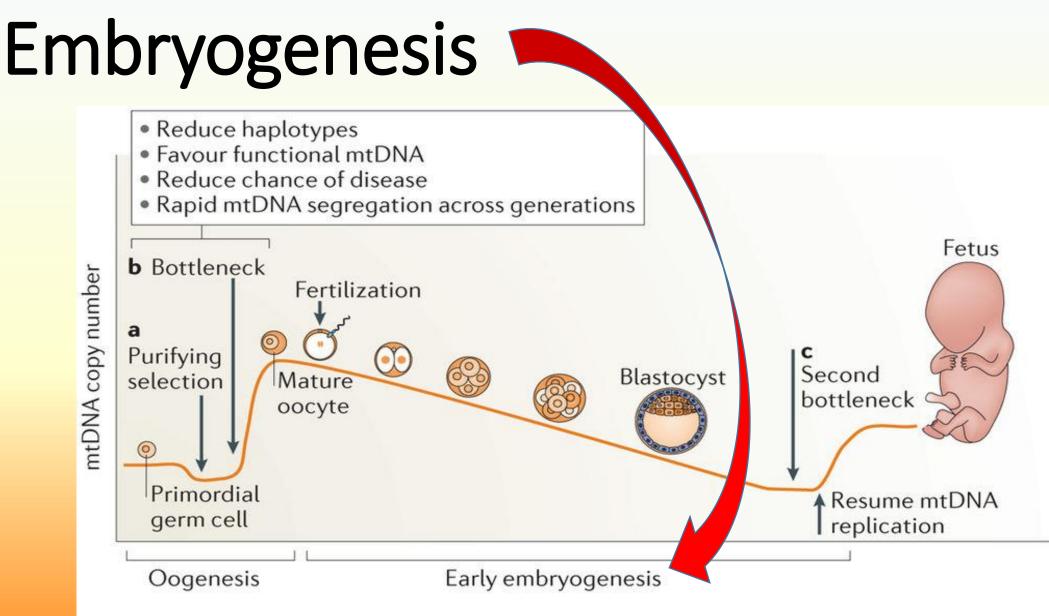
embryogenesis.

noun | em·bryo·gen·e·sis | _em-brē-ō-'je-nə-səs\

Popularity: Bottom 10% of words

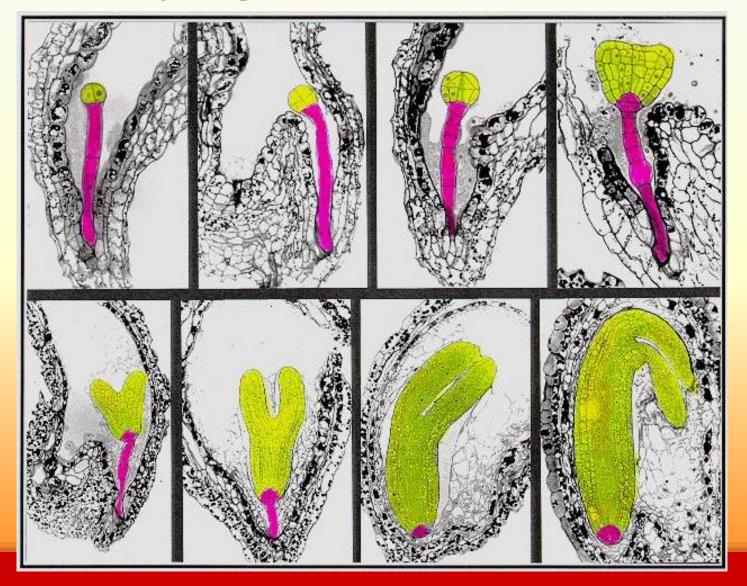
: the formation and development of the <u>embryo</u>



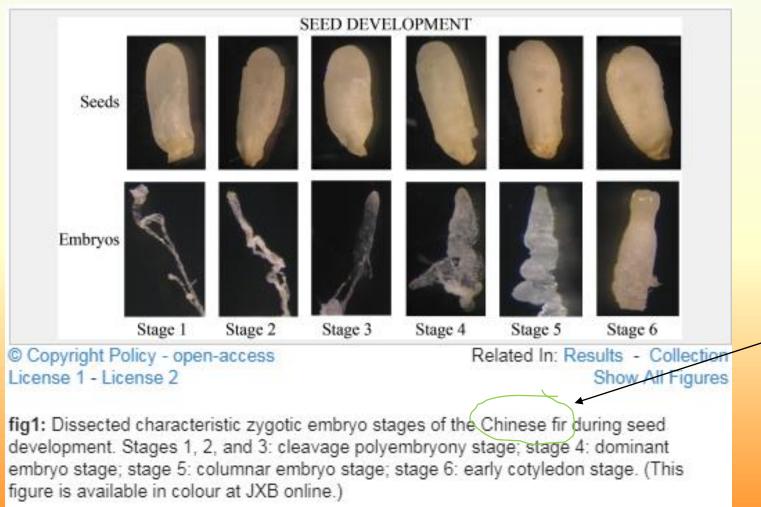


Nature Reviews | Molecular Cell Biology

Plant Embryogenesis



Zygotic Plant Embryogenesis



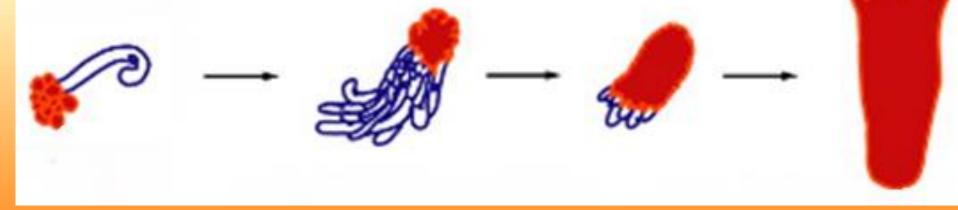
(Conifer example)

Zygotic Plant Embryogenesis

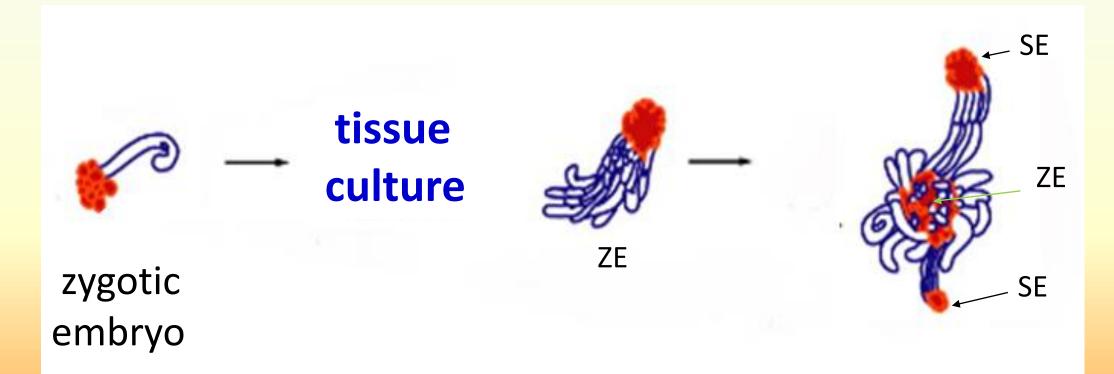


(Conifer example)

Inside a seed,.... Inside the megagametophyte



Somatic Plant Embryogenesis



Somatic Embryogenesis

SE

SE is often reported in conifers

<u>Genus</u>	<u>species</u>
001100	500000

- Abies 6
- Larix 6
- *Picea* 10

• other

• Pinus 15* * Klin

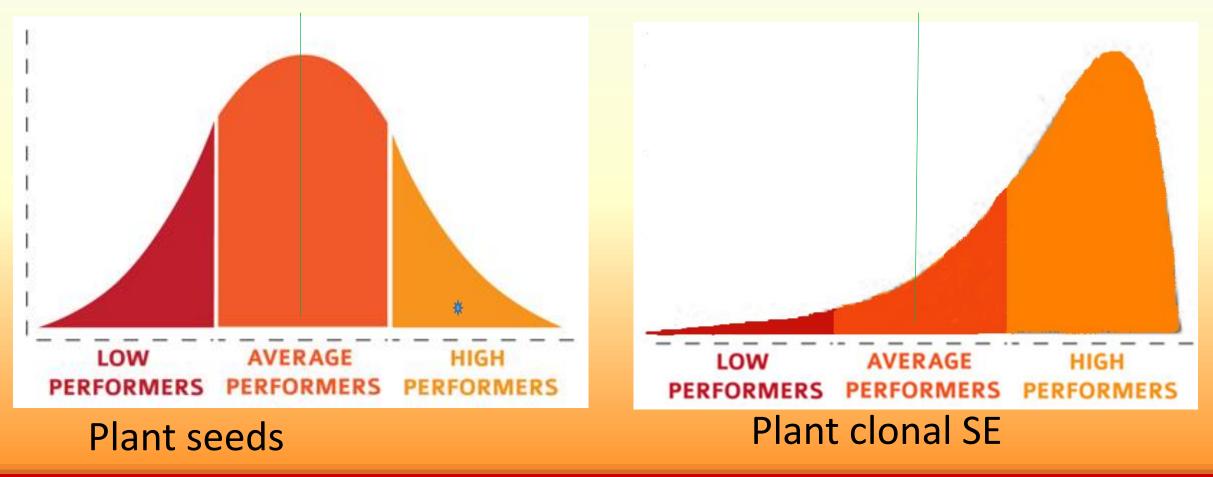
6

* Klimaszewska et al, Tree and Forestry Science and Biotechnology ©2007 Global Science Books

Claudio Stasolla, Lisheng Kong, Edward C. Yeung and Trevor A. Thorpe Source: In Vitro Cellular & Developmental Biology. Plant, Vol. 38, No. 2 (Mar. - Apr., 2002),pp. 93-105

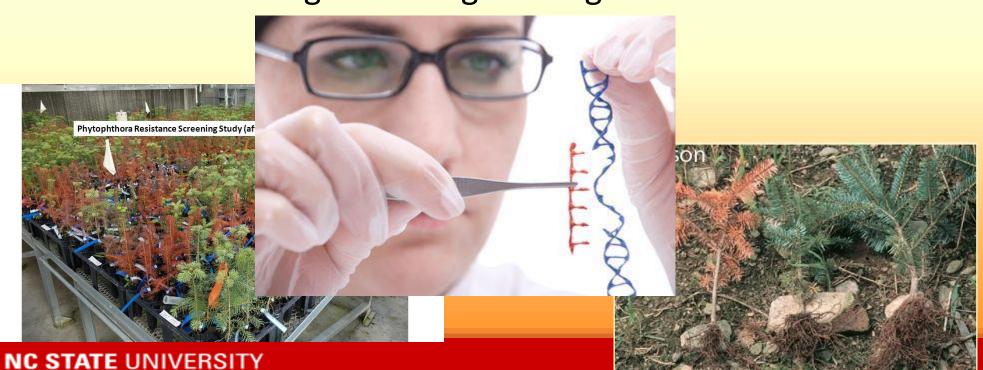
Why SE ?

• A method of cloning - make copies of one starting plant



Why SE ?

- Crop uniformity
- Embryos can be cryo-preserved for future use
- A vehicle for genetic engineering





Steps of the conifer SE process

Sterile *in vitro* culture

establish cultures of immature SE Initiation make millions of immature SE Proliferation cause the SE to mature into Maturation plantlets Acclimatization adapt the plantlets to soil and ex vitro conditions plant in field Deployment

Steps of the conifer SE process

<u>In vitro</u>culture

- Initiation
- Proliferation [cryopreservation]
- Maturation
- Acclimatization

• Deployment

establish cultures of immature SE make millions of immature SE

- cause the SE to mature into plantlets
- adapt the plantlets to soil and <u>ex vitro</u> conditions

plant in field

Molecular Tree Breeding lab

North Carolina State University

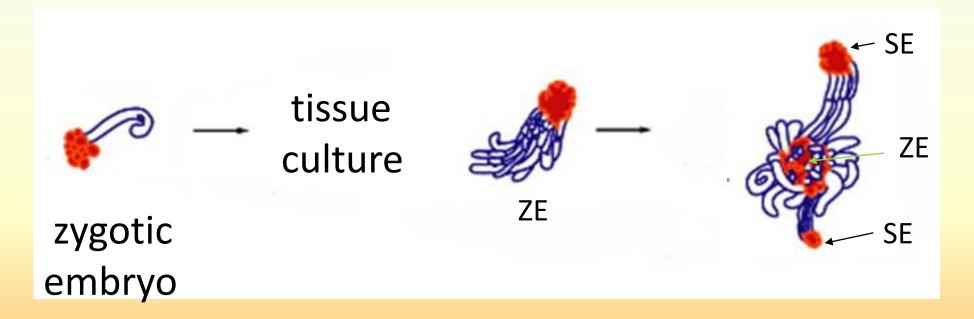


Frampton lab - Abies fraseri SE accomplishments

 Initiation Proliferation <pre>[cryopreservation]</pre> 	establish cultures of immature SE make millions of immature SE
 Maturation 	cause the SE to mature into plantlets
 Acclimatization 	adapt the plantlets to soil and <u>ex vitro</u> conditions
 Deployment 	plant in field

Initiation of SE cultures

Somatic Embryogenesis



SE Where does it begin?

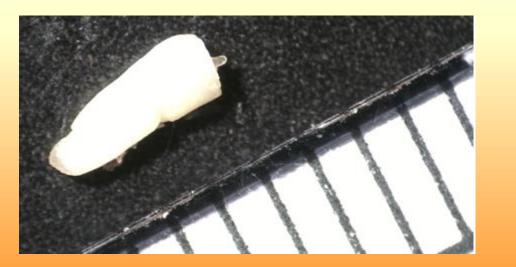
Mature zygotic embryo, stored 6 years, -20°C



5.0 mm

SE Where does it begin?

Immature zygotic embryo, ~ 30 post pollination



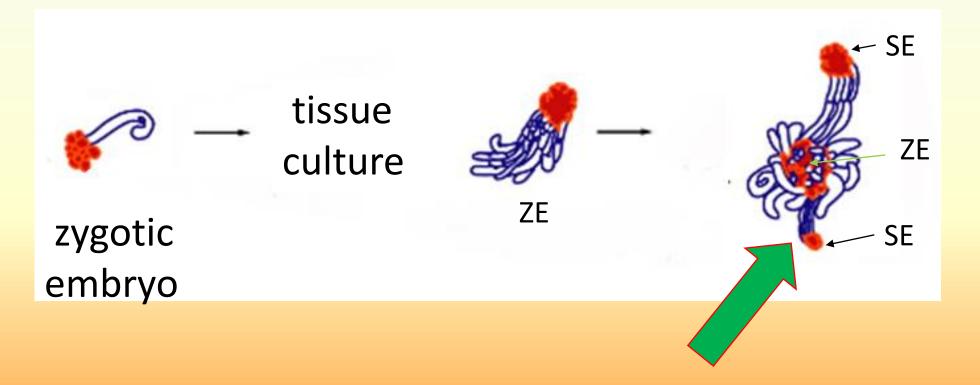


2016 SE Initiation Totals

		# embryogenic	
<u>zygotic embryo stage</u>	<u># processed</u>	<u>responses</u>	<u>%</u>
immature	4177	323	7.7
mature	1991	129	6.5

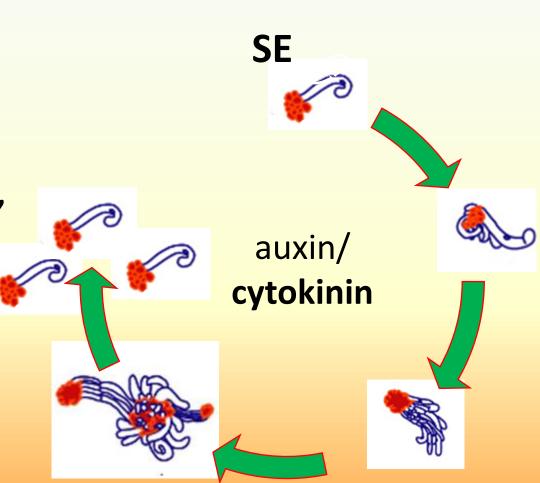
Proliferation of SE cultures

Somatic Embryogenesis



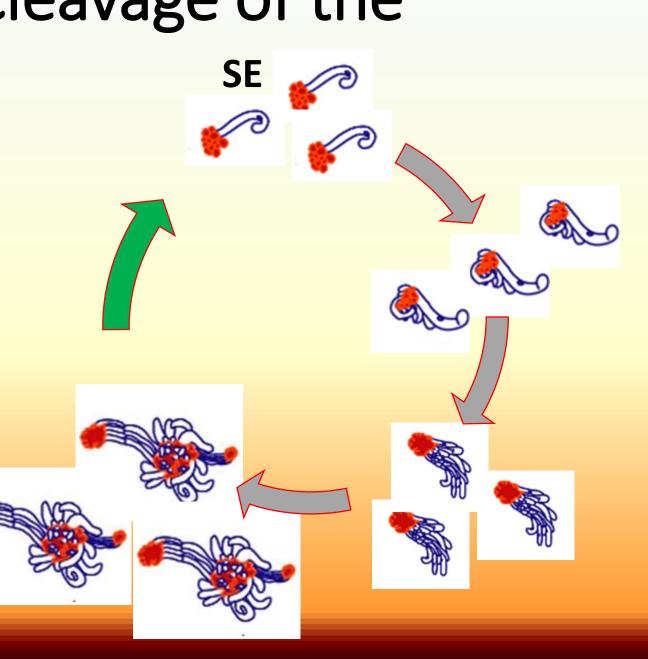
SE proliferate via cleavage of the embryo head se

- Most conifer SE cultures are proliferated using auxin:cytokinin, often in a 2:1 ratio.
- *Abies* SE cultures require **only cytokinin**



SE proliferate via cleavage of the embryo head

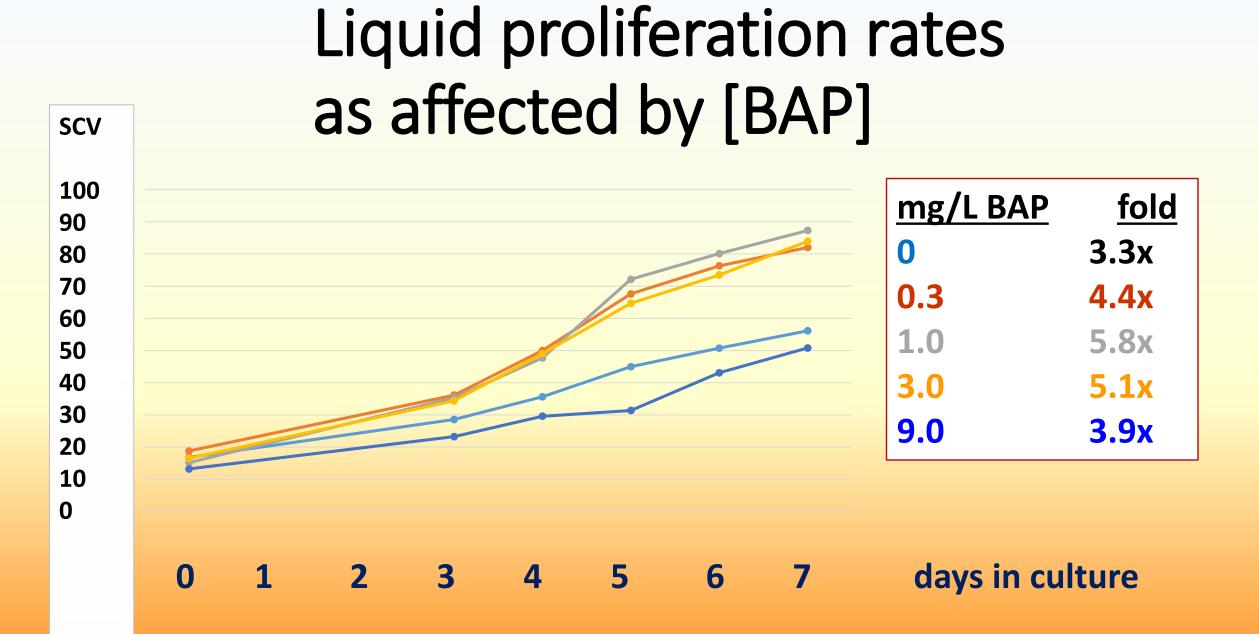
• Millions of copies from 1 starting embryo



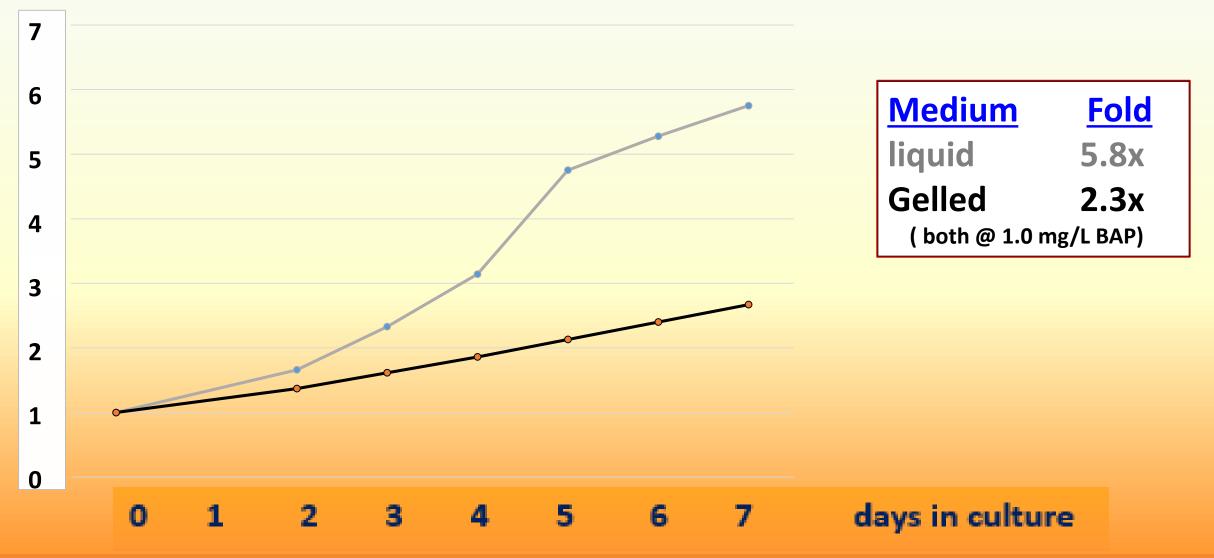
Proliferation with gelled or liquid media



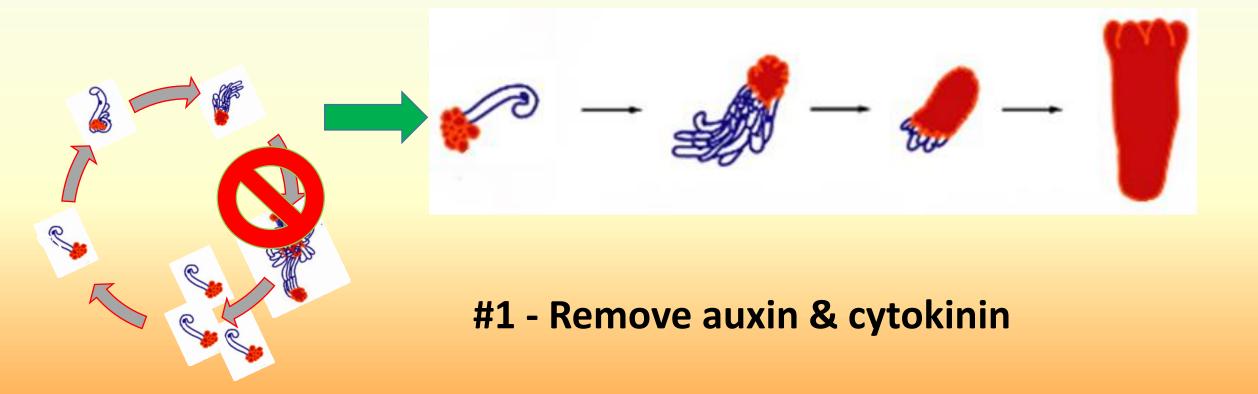




Liquid vs. gelled proliferation rates



SE Maturation



SE Maturation #2-4....

3 main components often used to achieve conifer SE maturation

ABA signal to accumulate storage products, developmental paths activated
Maltose possible developmental signal, osmoticant
PEG osmoticant, possible developmental signal

Concentrations of ABA, Maltose & PEG vary greatly across conifer species

			Abies			
	Pinus	Pinus	cilicica ×	Abies	Abies	Abies
species	caribaea	taeda	cephalonica	fraseri	alba	nordmanniana
year	2009	2003	2011	2016	2011	2002
mg/L ABA	31.7	5.3	10	5	5.3	10.6
g/L Maltose	64	20	40	40	40	45
g/L PEG	0	120	100	100	37.5	50

Design ABA, PEG & maltose "grids"

O Maltose	0 mg/L ABA	<mark>5</mark> mg/L ABA	10 mg/L AB	15 mg/L ABA
0 g/L PEG				
33 g/L PEG				
66 g/L PEG				
99 g/L PEG				

4 x 4 x 4 factorial = 64 combinations

	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/LABA
O g/LPEG			(GL	
33 g/L PEG	2	og/Lr	nal	0
66 g/L PEG			laltose	4
99 g/L PEG				Ó

	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/LABA
O g/L PEG		10		
33 g/L PEG		40 g∕L	m _{altose}	
66 g/L PEG			inditos(9
99 g/L PEG				

	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/LABA
O g/LPEG				
33 g/L PEG	•	<0 g/L	m _{a/tose}	
66 g/L PEG			. altose	
99 g/L PEG				

	<mark>0</mark> mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/L PEG		60		
33 g/L PEG		8/ ∠	m _{altos}	
66 g/L PEG			airos	e
99 g/L PEG				

Maturation test parameters

- Place ~ 200 mg tissue in a petri dish with 20 ml gelled medium
- 5 replicate plates per each of 64 treatments, and 1 control treatment
- Cultures are weighed at 0, 3, 6, 9 & 12 weeks
- Cotyledonary stage embryos are counted at weeks 6, 9 & 12.
- Tissue from 8 different genotypes tested

Cotyledonary Embryo Production

- Family Line # cot. E produced
- 11 33 0
 24 42 0
- 24 46 0
- 24 50 0
- 24 51 0
- 51 19 0
- 51 23 369
- 62 07 0

51-0023 average cotyledonary embryo / plate

O Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/LPEG				
33 g/L PEG				
66 g/L PEG				
99 g/L PEG				

40 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/L PEG				
33 g/L PEG	0.6	0.1	0.6	0.6
66 g/L PEG	12.9	1.9	0.1	0.8
99 g/L PEG	3.6			

20 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/L PEG				
33 g/L PEG			0.2	0.1
66 g/L PEG	1.7	0.1	1.0	
99 g/L PEG	3.0			

60 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/L PEG				0.1
33 g/L PEG	1.8	1		4.0
66 g/L PEG	10.0	2.0		
99 g/L PEG	0.5			

51-0023 total production - trends

- 0 maltose no embryo production
- Majority of embryos produced with 66 g/L PEG
- 40 g/L maltose > 60 g/L maltose
- Majority of embryos produced with 0 μM ABA



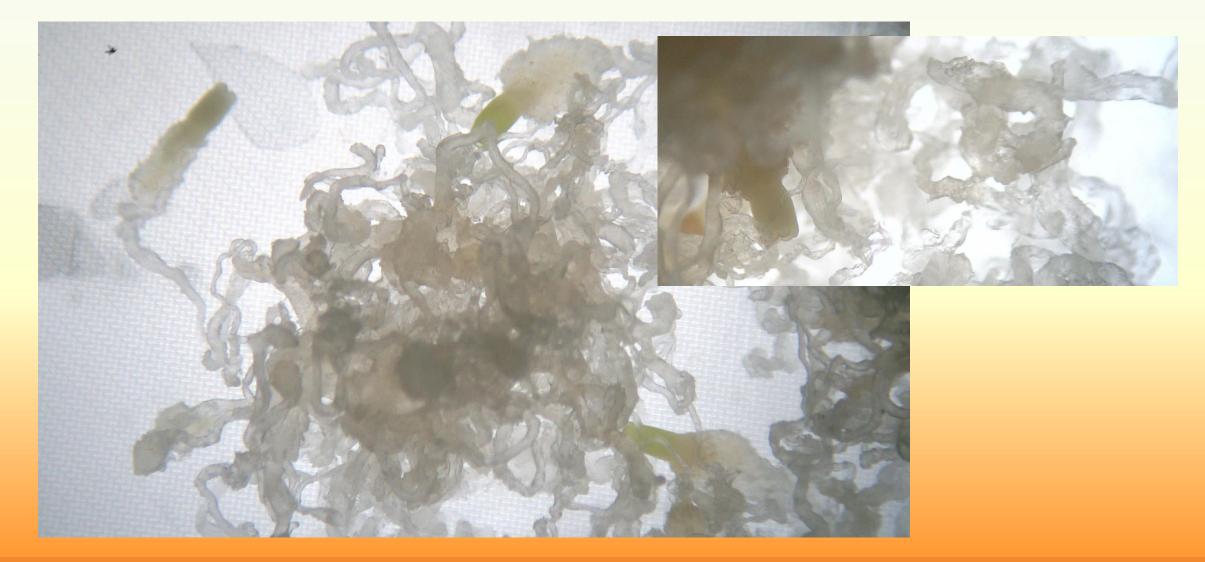
O Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/L PEG				
33 g/L PEG				
66 g/L PEG				
99 g/L PEG				
	0 g/L PEG 33 g/L PEG 66 g/L PEG		0 g/L PEG 33 g/L PEG 66 g/L PEG	0 g/L PEG 33 g/L PEG 66 g/L PEG

40 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/L PEG				
33 g/L PEG	0.6	0.1	0.6	0.6
66 g/L PEG	12.9	1.9	0.1	0.8
99 g/L PEG	3.6			

20 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/LPEG				
33 g/L PEG			0.2	0.1
66 g/L PEG	1.7	0.1	1.0	
99 g/L PEG	3.0			

60 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L ABA	15 mg/L ABA
O g/LPEG				0.1
33 g/L PEG	1.8	1		4.0
66 g/L PEG	10.0	2.0		
99 g/L PEG	0.5			

Fresh Weight Gain during Maturation



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Fresh Weight Gain during Maturation

in culture for 12 weeks, number shown = tota	I fold growth
--	---------------

O Maltose	0 mg/L ABA	<mark>5</mark> mg/L ABA	10 mg/L AB	15 mg/L ABA	40 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L AB	15 mg/L ABA
O g/LPEG	1.2	0.7	0.6	1.0	O g/L PEG	31.9	25	8.9	29.3
33 g/L PEG	.7	0.7	0.8	0.7	33 g/L PEG	29.9	28.2	30.1	25.2
66 g/L PEG	.7	0.7	0.7	0.8	66 g/L PEG	18.2	19.9	22.3	21.5
99 g/L PEG	.6	0.6	0.3	0.6	99 g/L PEG	42.3	6.8	9.4	6.5

20 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L AB	15 mg/L ABA	60 Maltose	0 mg/L ABA	<mark>5</mark> mg/L ABA	10 mg/L AB	. <mark>15</mark> mg/L ABA
O g/L PEG	14.9	11.7	10	8	O g/L PEG	45.9	51.8	44.9	36.5
33 g/L PEG	15.5	10.8	8.5	8.8	33 g/L PEG	49.8	28.2	27.7	21.1
66 g/L PEG	13.4	15.3	9.9	9.2	66 g/L PEG	13.1	15.3	6.9	6.3
99 g/L PEG	11.8	9.7	23.6	8.8	99 g/L PEG	12.9	5.6	2.7	2.7

control: 4.4 uM BAP 10 sucrose

O uM ABA **O** g/L PEG

5.6 x @ 12 weeks

(9.3 x @ 6 weeks)

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Next Steps.....

Yue Ken Liao Æ Cherng-Kang Liao Æ Ya Ling Ho

Plant Cell Tiss Organ Cult (2008) 93:257-268

species of *L. decidua* Mill., the embryogenic cultures will keep proliferating, rather than switching to a developmental phase, if the cultures contain a high amount of IAA (Korlach and Zoglauer 1995). It is

PHYSIOLOGIA PLANTARUM 116: 231–237. 2002 Copyright C Physiologia Plantarum 2002 Printed in Denmark – all rights reserved ISSN 0031-9317

Effect of anti-auxins on maturation of embryogenic tissue

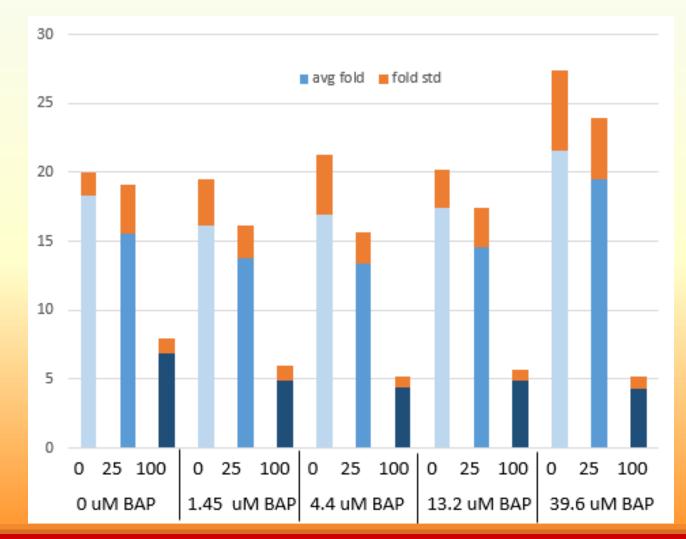
cultures of

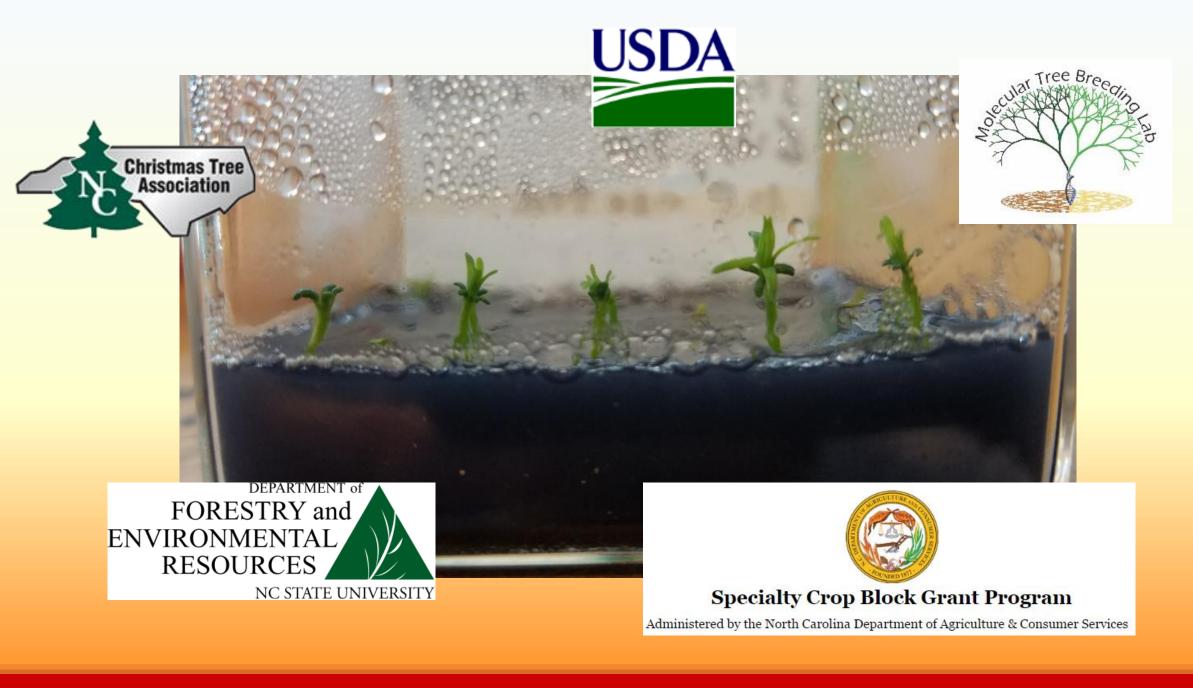
Nordmanns fir (Abies nordmanniana)

Inclusion of PCIB into the maturation medium had two general effects: (1) proliferation was reduced, and (2) the number of embryos that converted from proliferation to maturation was significantly increased. However, both

PCIB slows Abies fraseri proliferation

UM PCIB 0 25 100





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People in Denmark is currently working in SE in Normand fir!!!!!!



They have 400 clones total in 6 trials – 8000 ramets in total planted autumn 2014 and 2015

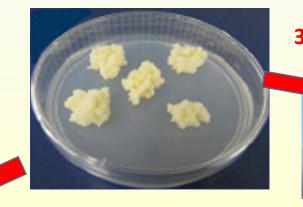


Ideal Fraser fir SE Process

1) Pick your favorite tree



2) Establish tissue cultures



3) Cryopreserve sample

4 & 5) Convert embryos into seedlings







6) Fill greenhouse with 1000's of clones, <u>exact copies</u> of your favorite tree

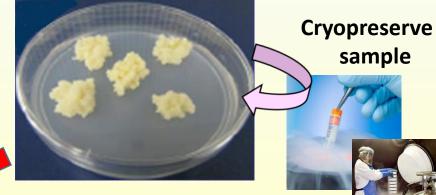


Ideal Fraser fir SE Process

1) Select an elite tree



- 2) Initiate embryogenic tissue culture
- 3) Multiple embryos



4) Mature embryos into seedlings



5) Acclimatize in greenhouse



6) Plant 1000's of copies of your favorite tree

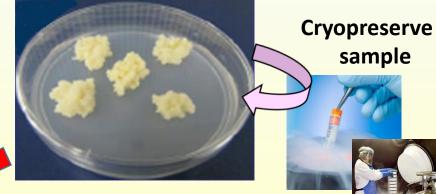


Ideal Fraser fir SE Process

1) Select your favorite tree



- 2) Initiate embryogenic tissue culture
- 3) Multiple embryos



4) Mature embryos into seedlings



5) Acclimatize in greenhouse



6) Plant 1000's of copies of your favorite tree



1) Select your favorite trees



Maturation weight trends

Increasing maltose increases F.W.T. gain

Increasing PEG decreases F.W.T. gain

O Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L AB	15 mg/L ABA
O g/LPEG	1.2	0.7	0.6	1.0
33 g/L PEG	.7	0.7	0.8	0.7
66 g/L PEG	.7	0.7	0.7	0.8
99 g/L PEG	.6	0.6	0.3	0.6

20 Maltose	0 mg/L ABA	5 mg/L ABA	10 mg/L AB	15 mg/L ABA
O g/L PEG	14.9	11.7	10	8
33 g/L PEG	15.5	10.8	8.5	8.8
66 g/L PEG	13.4	15.3	9.9	9.2
99 g/L PEG	11.8	9.7	23.6	8.8

40 Maltose	0 mg/L ABA	<mark>5</mark> mg/L ABA	10 mg/L AB	15 mg/L ABA
O g/LPEG	31.9	25	8.9	29.3
33 g/L PEG	29.9	28.2	30.1	25.2
66 g/L PEG	18.2	19.9	22.3	21.5
99 g/L PEG	42.3	6.8	9.4	6.5

60 Maltose	<mark>0</mark> mg/L ABA	5 mg/L ABA	10 mg/L AB	15 mg/L ABA
O g/L PEG	45.9	51.8	44.9	36.5
33 g/L PEG	49.8	28.2	27.7	21.1
66 g/L PEG	13.1	15.3	6.9	6.3
99 g/L PEG	12.9	5.6	2.7	2.7

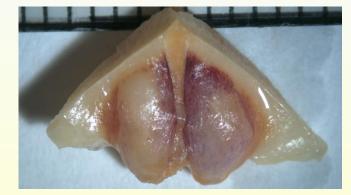
0 uM ABA 0 g/L PEG ucrose

5.6 x @ 12 weeks

(9.3 x @ 6 weeks)

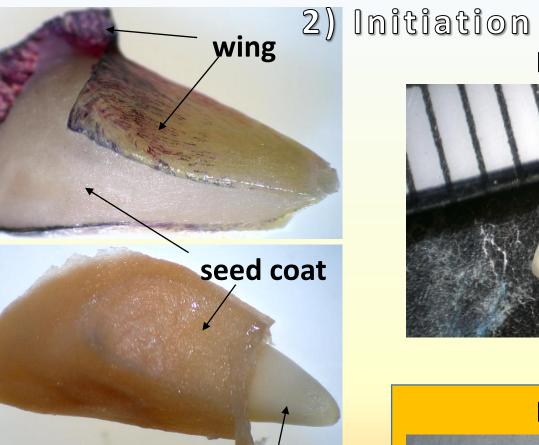


Immature embryos, 2-4 weeks/year

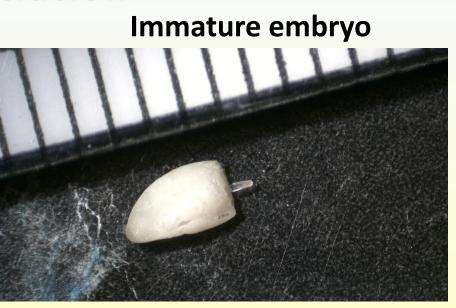


Mature embryos year round





megagametophyte



Mature embryo



2) Initiation +

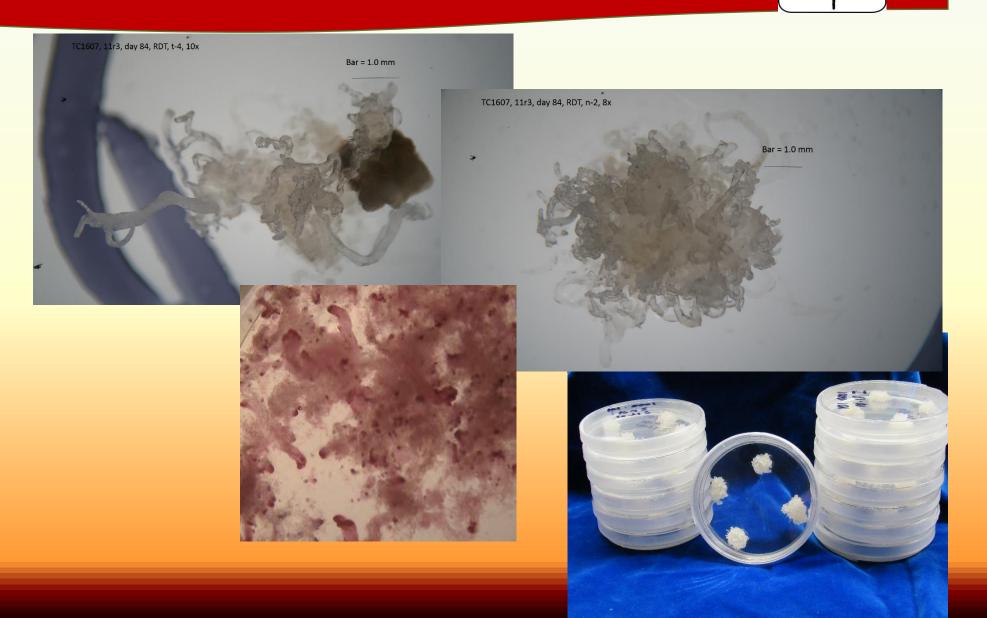
Mature embryo



Immature embryo



3) Multiplication

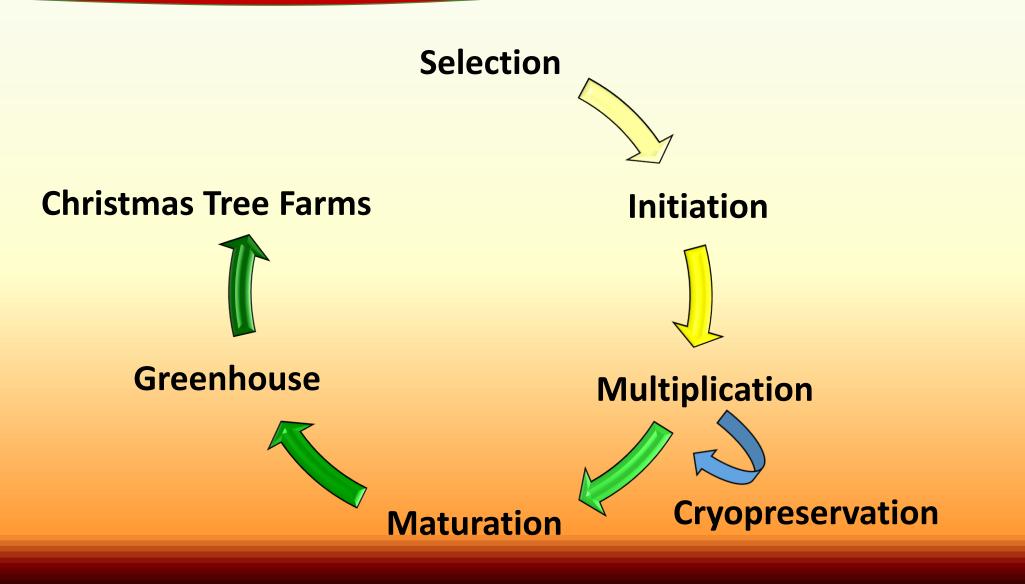


3) Multiplication





Somatic Embryogenesis Process Flow

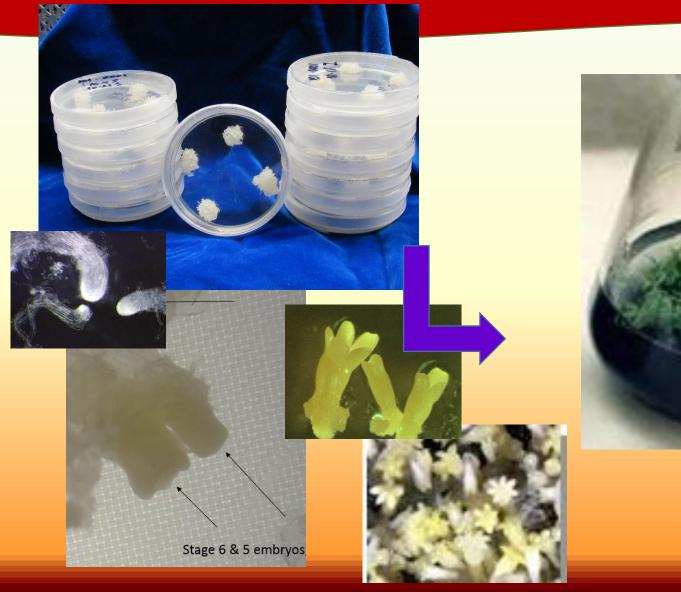






4) Maturation





5) Acclimatization









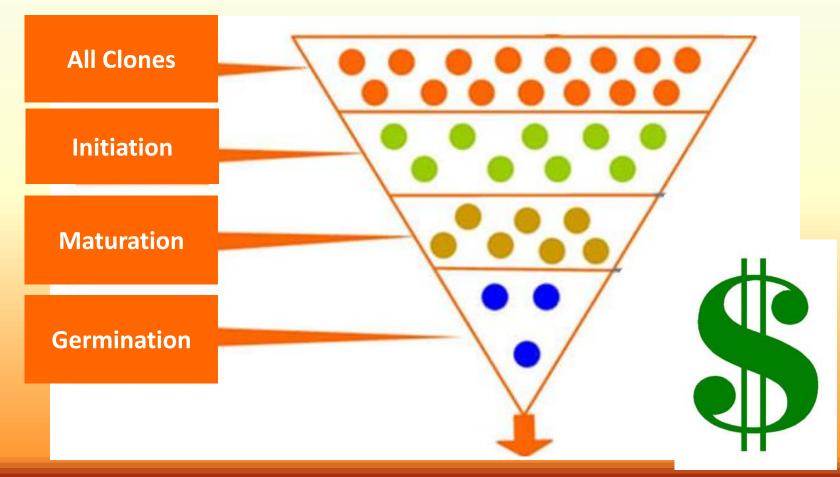
Plant a Christmas Tree Farm



Current Challenges



Techniques work on only some clones



Controlled Pollination

1) Clonal Selection



Embryo Rescue





Abies fraseri

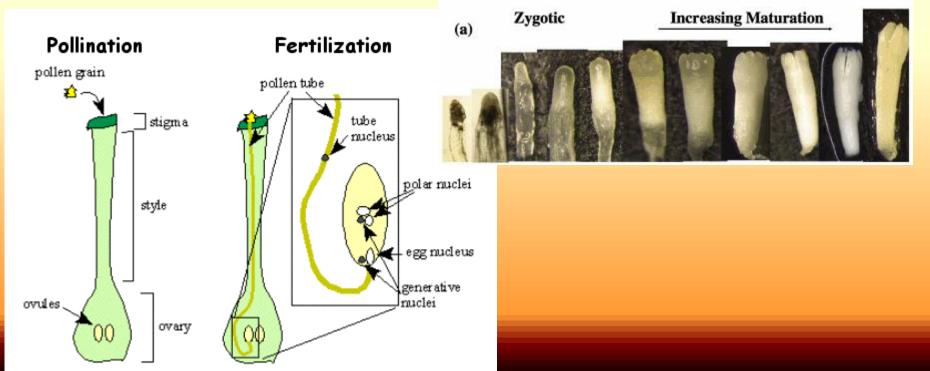
Abies firma

Cross 2 different species & get ~no viable seed

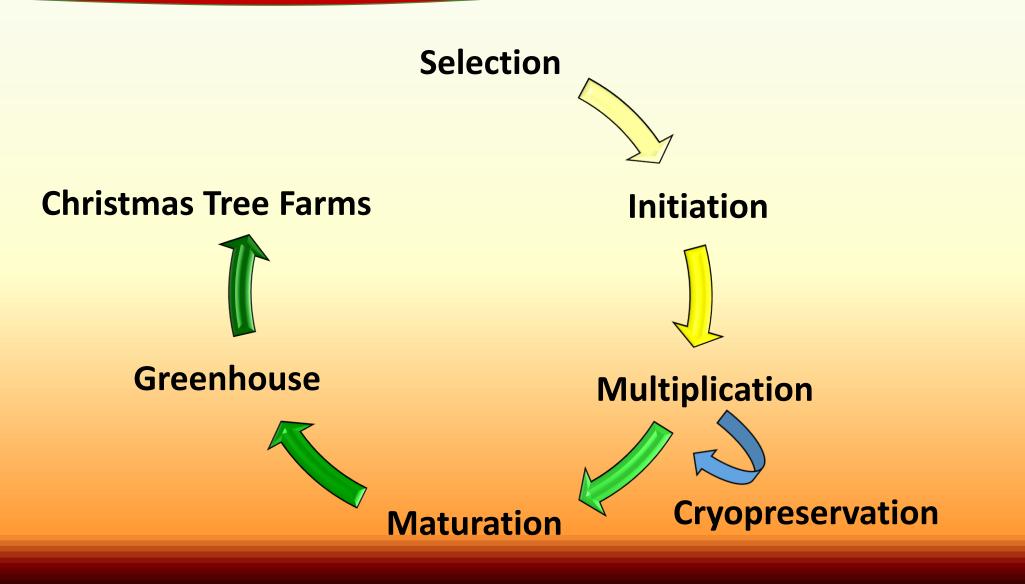
Interspecific hybrids

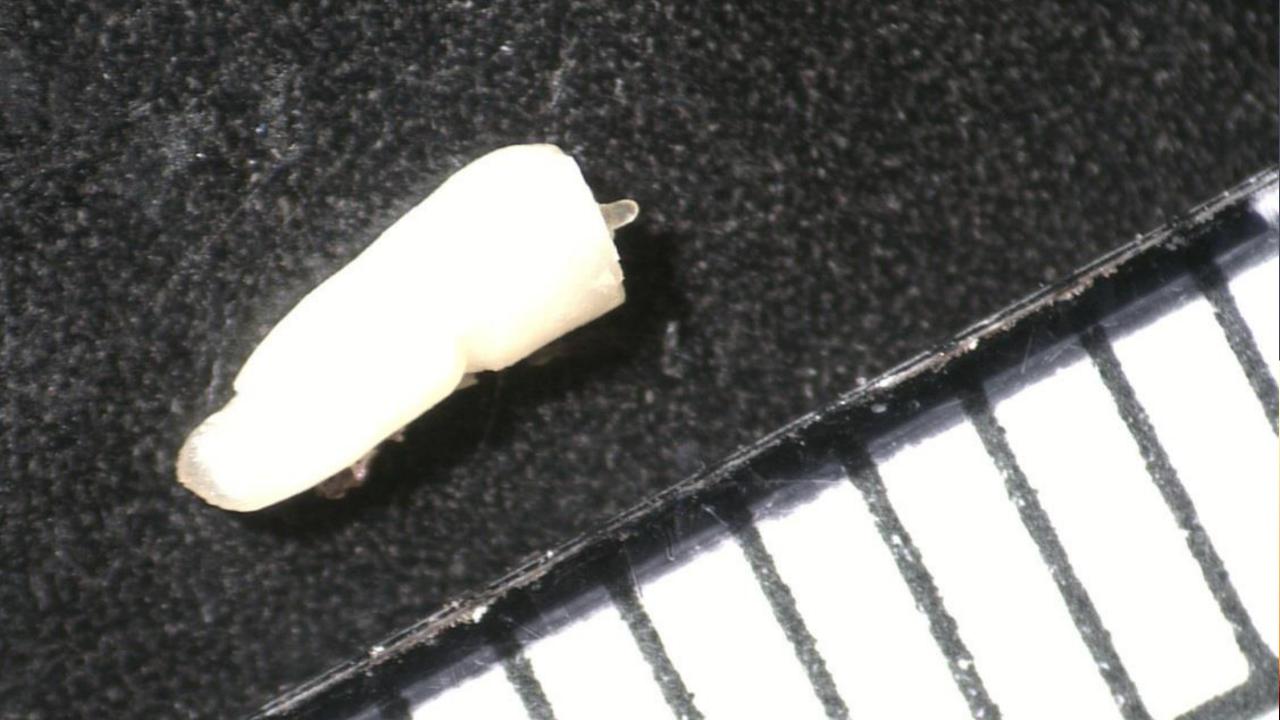


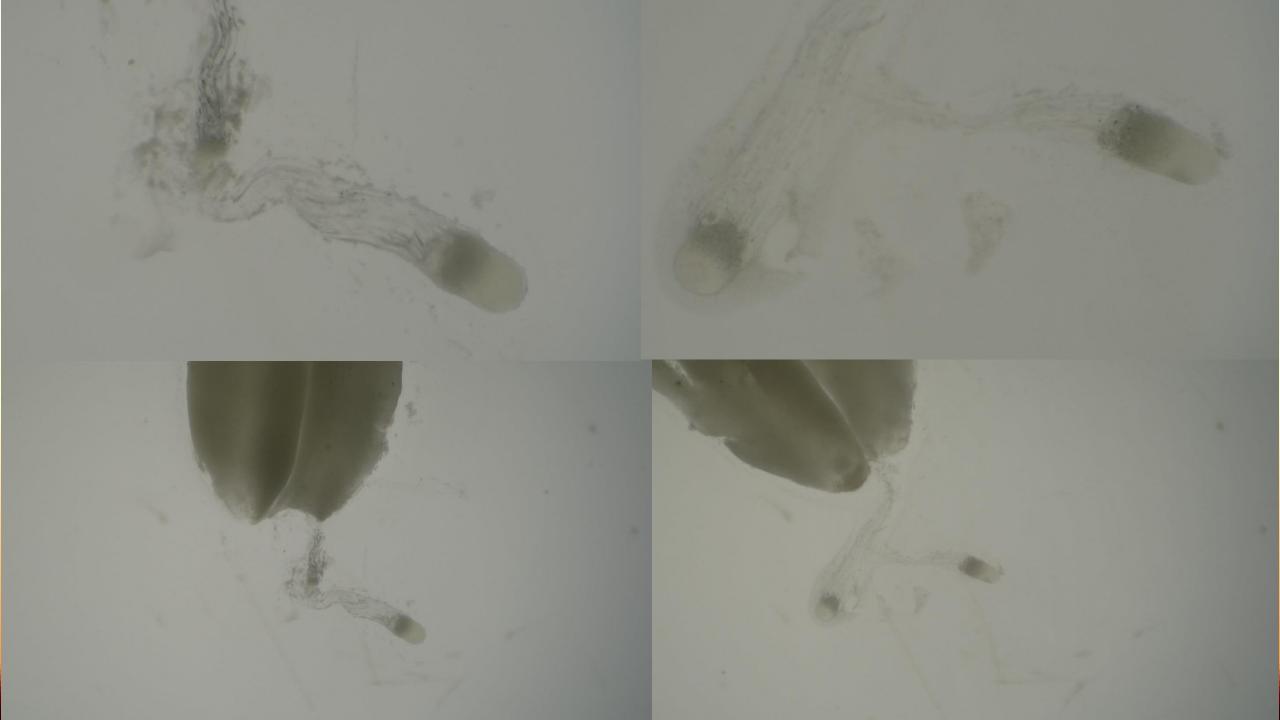
Interspecific hybrids are bred by <u>mating two</u> <u>species</u>, normally from within the same genus. The <u>offspring display traits of **both** parents</u>, but <u>are often sterile</u>.



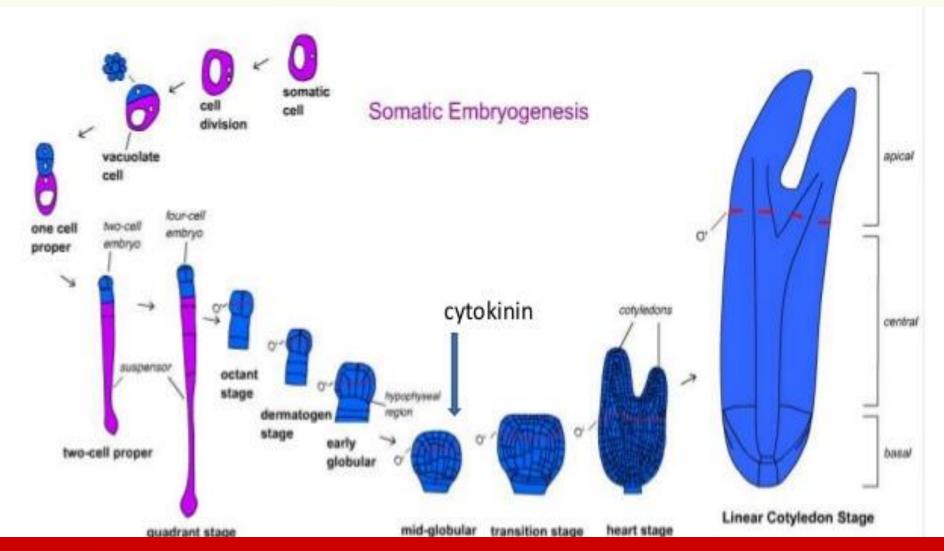
Somatic Embryogenesis Process Flow







Stages of Somatic Embryogenesis

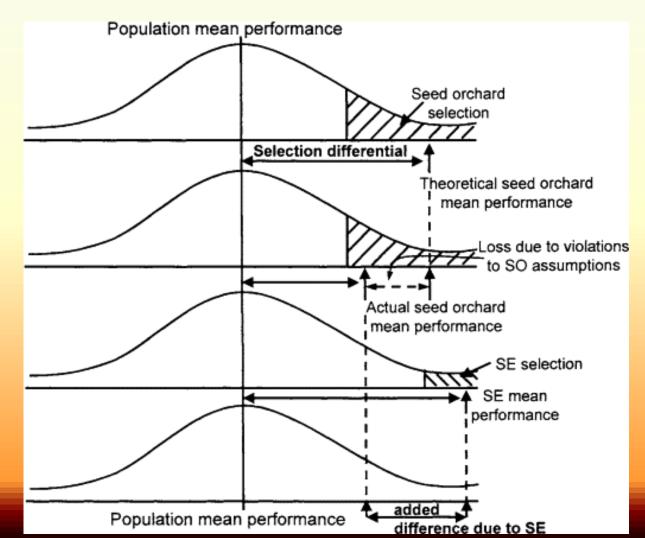


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Why SE ?

• A method of cloning - make copies of a starting plant



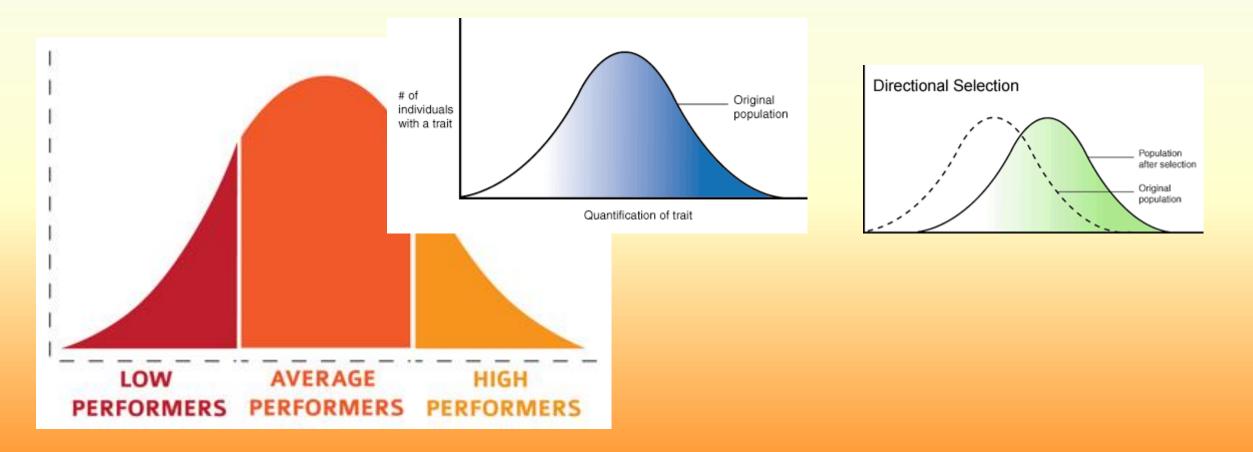
Increased Productivity

SE Process Flow

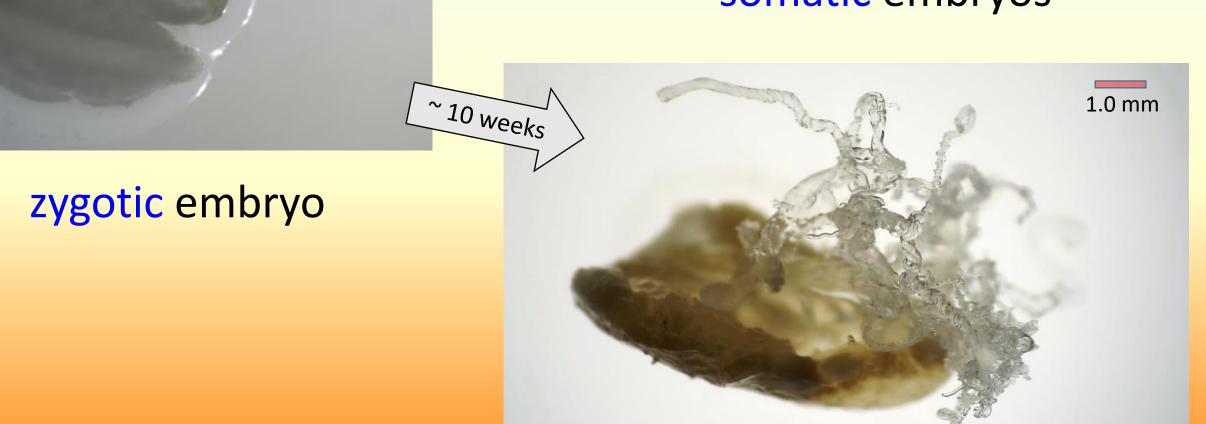
- Initiate SE from zygotic embryos
- Proliferate the few immature SE up to millions/line
- Mature the SE into cotyledonary embryos with roots
- Acclimate the SE to soil/greenhouse coniditons
- Deploy to field

Why SE ?

• A method of cloning - make copies of one starting plant



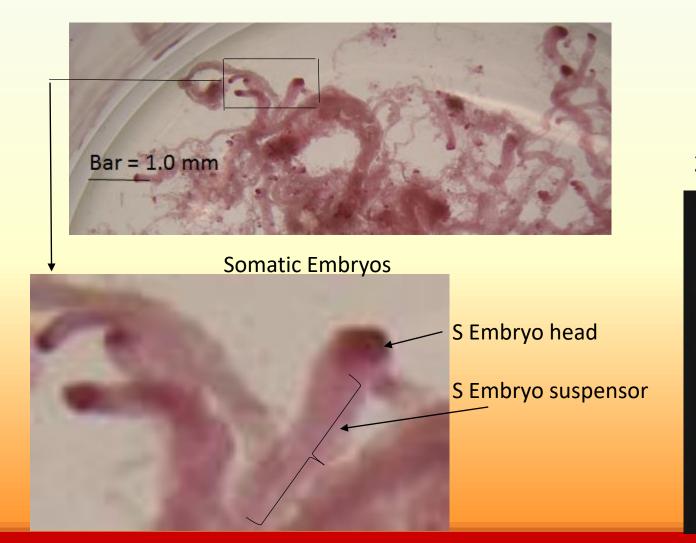
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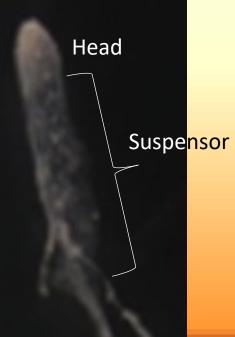
1.0 mm

proliferating somatic embryos

SE stained with acetocarmine



Zygotic Embryo



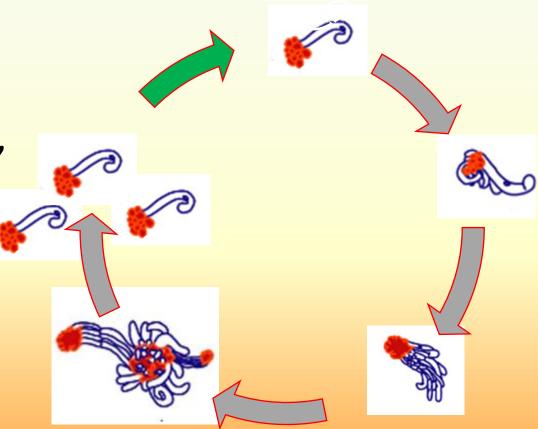
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Additional advantages of suspension culturing

- uniform exposure of tissue to any chemical variable being tested
- uniform tissue inoculum when aliquoting tissue to experiments
- general ease of handling
- reduces plastic waste petri dishes

SE proliferate via cleavage of the embryo head

- Most conifer SE cultures are proliferated using auxin:cytokinin, often in a 2:1 ratio.
- *Abies* SE cultures require **only cytokinin**



SE proliferate via cleavage of the embryo head

